# **Advanced Technology and Biology Division WEHI Proteomics Facility**



Established in 2021, we provide the latest mass spectrometric instrumentation for researcher-led exploration, identification and quantification of proteins and peptides. We are a team of 6 postdoctoral scientists specialising in mass spectrometry-based proteomics.

From experimental design and sample preparation, to data analysis and statistical evaluation, our staff can undertake a broad range of proteomics experiments on behalf of users which are cost-recovered at a WEHI-subsidised hourly rate.

Between 2021-2023, we have processed >12,000 samples that were submitted by 90 collaborating labs (55 internal, 25 external) across 543 projects with an average completion time within 60 days of sample submission. We have overseen 106 development projects towards expanding proteomics workflows, totalling >3500 hours.



#### **Current research projects / research interests**

2024 development (and beyond!) SDA-MS (structural proteomics) Laser-capture microscopy proteomics Phosphoproteomics (with Tanzer Lab) Antibody characterisation

Computational proteomics Proteome normalization Missingness & imputation Batch correction Machine-learning models At WEHI Proteomics, we offer a wide range of workflows that have been refined and optimised via multiple in-house research projects driving our *methods development*.





These are reliant on our *mass spectrometry* technology platform consisting of high resolution LC-MS instrumentation.

Since 2021 we have included computational proteomics as part of our services, which has accelerated our research in proteomicsbased statistics and data analysis.

Notably, WEHI Proteomics has maintained strong track record in *clinical biomarker* discovery towards fast-tracking translation.

# Technologies, techniques, and instrumentation



# **Recent Top Publications**

1. Meng, Y., Garnish, S.E., Davies, K.A. et al. *Phosphorylation*dependent pseudokinase domain dimerization drives fulllength MLKL oligomerization. Nat Commun 14, 6804 (2023).

2. Low, R.R.J., Fung, K.Y., Gao, H. et al. S100 family proteins are linked to organoid morphology and EMT in pancreatic cancer. Cell Death Differ 30, 1155–1165 (2023).

3. Triglia, T., Scally, S.W., Seager, B.A. et al. *Plasmepsin X* activates the PCRCR complex of Plasmodium falciparum by processing PfRh5 for erythrocyte invasion. Nat Commun 14, 2219 (2023).

4. Roy, M.J., Surudoi, M.G., Kropp, A. et al. Structural mapping of PEAK pseudokinase interactions identifies 14-3-3 as a molecular switch for PEAK3 signaling. Nat Commun 14, 3542 (2023)

Antibody *de novo* sequencing Antibody characterisation Epitope mapping

Low-input/LCM Proteomics Low-input, high-sensitivity Laser-capture microscopy samples

(Bruker)

(Bruker)

(Thermo Fisher Scientific)

### **Services offered by WEHI Proteomics**



**1. MS sample preparation** 

WEHI Proteomics staff prepare samples for proteomics analysis. Users need only provide cell pellets, biofluids (e.g. plasma/urine), or cell/tissue lysates.

Our services include: Protein quantitation (e.g. BCA), sample enrichment (e.g. streptavidin affinity purification, exosome enrichment), C18 cleanup/peptide desalting, phosphopeptide and diGly enrichment.

Users can also be trained, or perform sample preparation supervised by facility staff.

### **Recent achievements**



Since 2021, we have increased our capacity across the board resulting in an almost doubling of project submissions, collaborators and samples!



2. MS instrument acquisition

Samples are queued, injected and analysed on one of our 7 mass spectrometers via liquid chromatography and MS spectra are acquired.

Our MS instruments include: 2 x timsTOF Pro MS (Bruker) 1 x Orbitrap Astral MS (Thermo) 1 x Orbitrap Eclipse Tribrid MS (Thermo) 1 x Q-Exactive Classic MS (Thermo) 1 x maXis II Q-TOF MS (Bruker) (Native MS) 1 x Impact II Q-TOF MS (Bruker) (Intact MS)



#### 3. Proteomics data analysis

We have an in-house biostatistician (Dr Jumana Yousef) who assesses all data and undertakes data QC and statistical analyses (e.g. differential expression).

*Our routine data analysis includes:* R markdown .html document which contains all steps performed in the analysis • Interactive Spotfire document of all results

We also undertake additional analyses upon request including time-course analysis, kmeans clustering, correlation analysis, GO enrichment and more!

WEHI Proteomics charges an hourly rate for time spent by facility staff on: 1) sample preparation 2) MS instrument time 3) data analysis

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Low Input/

GPF

LCM

Proteomics

Development

Our hourly rate is subsidised by WEHI for all internal researchers.

Note, we don't charge for pre-project meetings, **but** we encourage them!

Users should reach out to discuss future experiments before they begin collecting samples so we can help with experimental design and ensure the best results!



With our staff dedicating 0.2 FTE towards method

## **Meet the team**









Dr. Laura Dagley: Facility manager BBiomedSc (Hons) *Melbourne*, PhD *Melbourne* 

Dr. Samantha Emery-Corbin: Deputy facility manager & clinical proteomics BSc (Hons), PhD *Macquarie University* 

Dr. Toby Dite: Structural, Intact and PTM Specialist BBiomedSc (Hons) *Melbourne*, PhD *Melbourne* 

Dr. Dylan Multari: General Proteomics MRes, PhD *Macquarie University* 

Dr. Jumana Yousef: Bioinformatician/Statistician Statistics (MSc) NMSU, Bioinformatics (MSc) Melbourne, PhD Monash

**Dr. Vineet Vaibhav: Clinical and Cell Surface proteomics** BSc BHU, Biotechnology (MSc) IIT Bombay, PhD Macquarie University



Due to method development improvements, we

halved MS instrument times per sample, reducing



**2021:** 139 **2021:** 34 2021: 3261 **2022:** 177 **▲** 27% **2022:** 48 ▲ 41% **2022:** 4474 **▲** 37% **2023:** 49 **▲** 44% **2023:** 4873 **▲** 49% **2023:** 217 ▲ 56%



development and protocol optimisations, we have several new methods available within our facility!



